





APLICOR 3D

Procedure Checklist

Personalized power, realized now.

MKT-197, Rev. 02

supply checklist

SUPPLIES PROVIDED

Items	Qty*
Matrix plate	1
Micronizers (variety)	4
Dispenser 1	1
PCL • Small Kit: 364mm • Medium Kit: 667mm • Large Kit: 1,100mm	1
Luer connector	1-2
NFC tag	1
Sensor Tray	1
Bed film	1



ADDITIONAL SUPPLIES (not provided)

Items	Qty
Waste tray	1
Standing rack	1
Normal saline (50-100ml)	1
Syringe caps	2
21-gauge, 1 inch needle*	2
5 ml syringes*	2-4
10ml syringes	2-4
50ml syringes	1-2
Surgical tools	N/A

^{*}These supplies <u>must be the exact</u> <u>specifications</u>. No substitutions. Additional supplies without asterisks can be substituted at the professionals' discretion.



^{*} Quantity based on size of kit

	Operation		
Pra	ctice Steps	Role	✓
1	Wound Bed Preparation		
ı	a. Perform debridement to clean and prepare the wound bed	S	
	Generation and Transfer of Regen File to APLICOR 3D Printer using AiD Regen Tablet		
	a. Tap [Start] at the home screen \rightarrow Select the surgeon from the list	1	
	b. Enter the patient code or register a new patient $ ightarrow$ Select the wound type	1	
	c. Add photos of the wound (snap a photo or bring a photo from the gallery)	1	
2	d. Select the wound area by Al-assisted detection or by manual drawing	1	
	e. Set the patch thickness and printing type	1	
	f. Request a Regen file	1	
	g. Enter the IP address shown on the LCD screen of APLICOR 3D Printer	1	
	h. Send the Regen file to APLICOR 3D Printer	1	
	i. Check the required volume of fat harvesting and each bioink	1	
	※ Step 3 & Step 4 can be done simultaneously or vice versa		
	Scaffold Printing		
	a. Tap [Confirm] on APLICOR 3D Printer LCD screen	1	
	b. Open the door of APLICOR 3D Printer and Insert the NFC Tag	1	
	c. Insert NFC Tag and Sensor Tray	2	
	d. Check print information → Tap the file	1	
	e. Tap [Scaffold]	1	
3	f. Insert Matrix Plate coated with Bed Film	2	
	g. Tap [OK] to proceed	1	
	h. Insert Dispenser 1 into Dispenser 1 Dock	2	
	I. Install 3D Regen PCL and load it into the inlet of Dispenser 1 Dock	2	
	j. Close the door of APLICOR 3D Printer	1	
	k. Perform printing (which will start once the closed door is detected)	1	
	I. Check the printed Scaffold	1	
4	Fat Harvesting		

operation checklist

S surgeon 1 non-sterile staff 2 surgeon or sterile staff

	Operation		
Pra	ctice Steps	Role	✓
	Adipose Tissue Micronization		
_	 a. Bring the harvested adipose tissue to the sterile work area where the APLICOR 3D Scaffold Kit components are located 	2	
	b. If the harvested fat is contained in a 20~50ml syringe, aliquot the fat into separate 10ml luer lock syringes using luer-to-luer Connector	2	
5	c. Micronize the adipose tissue with Adipose Micronizer ①	2	
	d. Micronize the adipose tissue with Adipose Micronizer ②	2	
	e. Micronize the adipose tissue with Adipose Micronizer ③	2	
	f. Micronize the adipose tissue with Adipose Micronizer ④	2	
	Washing of Micronized Adipose Tissue with Normal Saline		
	a. Introduce the micronized adipose tissue to sterile normal saline contained in a 30ml or 50ml luer lock syringe using luer-to-luer Connector (Recommended ratio - Micronized adipose tissue : normal saline = 1:2 ~ 1:5)	2	
,	b. Close the syringe containing both the adipose tissue and the normal saline with a needle or syringe cap	2	
6	c. Gently mix by inverting	2	
	d. Place the syringe on a standing rack with the needle or syringe cap facing down	2	
	e. Wait until sufficient layer separation has been achieved	2	
	f. Discard the blood and fluid layers beneath the adipose layer into the waste tray	2	
	g. Obtain the final micronized and washed adipose tissue (i.e., ECM)	2	
	Bioink Preparation		
	a. In case of Fibrin Protocol:	2	
	a) Check AiD Regen App to determine the required volume for each Bioinks [Bioink 1= ECM & Fibrinogen / Bioink 2 = Thrombin]	2	
	b) Prepare the required volume of Fibrinogen in an empty luer lock syringe	2	
	c) Prepare the required volume of ECM in an empty luer lock syringe	2	
7	d) Prepare Bioink 1 by gently mixing the prepared ECM and Fibrinogen at the proper ratio using Dispenser 2 Tube	2	
	e) Transfer Bioink 1 into 5ml luer lock syringe using luer-to-luer connector	2	
	f) Close 5ml luer lock syringe containing Bioink 1 with 21G 1-inch needle	2	
	g) Transfer the required volume of Thrombin (Bioink 2) into 5ml luer lock syringe (luer-to-luer connector or 3-way IV connector can be used)	2	
	h) Close 5ml luer lock syringe containing Bioink 2 with 21G 1-inch needle	2	

	Operation			
Pra	ctice Steps	Role	✓	
7	b. In case of Low Temperature Protocol:			
	a) Check AiD Regen App to determine the required volume for ECM	2		
	b) Prepare the required volume of ECM in 5ml luer lock syringe using luer-to- luer connector	2		
	c) Close 5ml luer lock syringe containing Bioink 1 with a 21G 1-inch needle	2		
	Bioink 1 & Bioink 2 Printing using APLICOR 3D Printer			
	a. Tap [Bioink 1] on the LCD screen of APLICOR 3D Printer	1		
	b. Open the door of APLICOR 3D Printer	1		
	c. Remove the cap of the needle on 5ml Luer lock syringe containing Bioink 1	2		
	d. Load 5ml Luer lock syringe containing Bioink 1 into Dispenser 2 Dock	2		
	e. Close the door of APLICOR 3D Printer	1		
	f. Perform printing (which will start once the closed door is detected)	1		
	g. When printing is complete, open the door.	1		
8	g. Remove 5ml Luer lock syringe from Dispenser 2 Dock	2		
	*Following steps (from h to n) are only for Fibrin Protocol			
	h. Tap [Bioink 2] on the LCD screen of APLICOR 3D Printer	1		
	i. Remove the cap of the needle on 5ml Luer lock syringe containing Bioink 2	2		
	j. Load 5ml Luer lock syringe containing Bioink 2 into Dispenser 2 Dock	2		
	k. Close the door of APLICOR 3D Printer	1		
	I. Perform printing (which will start once the closed door is detected)	1		
	m. When printing is complete, open the door.	1		
	n. Remove 5ml Luer lock syringe from Dispenser 2 Dock	2		
	Solidification			
	a. In case of Fibrin Protocol :			
9	a) Wait for 20~30 minutes (the elapsed time appears on the LCD screen)	2		
	b) After 15~20 minutes, gently scrape the sides of the patch against the edge of the PCL scaffold	2		
	c) Gently scrape the patch off the Bed Film	2		

	Operation		
Pra	ctice Steps	Role	✓
	b. In case of Low Temperature Protocol:		
	a) Wait for 10~15 minutes (Keep the door of APLICOR 3D Printer closed)	2	
	b) After 10~15 minutes, check the patch solidity by touching it with Dispenser 2 Tip	2	
	c) Cut one side of the Scaffold with a scalpel	2	
9	d) Peel off the Scaffold with forceps or with hands	2	
	e) Separate the matrix plate from the plate bed and check whether the patch is separated from the bed film using forceps after several seconds.	2	
	f) If the patch is separated from the bed film, put the Matrix Plate back on the plate bed and store it at a low temperature until applied to the wound	2	
	Patch Application to the Wound		
	a. Pick up the patch with forceps	2	
10	b. Apply the patch to the wound site	2	
	c. Apply the first dressing	2	
	d. Apply the second dressing	2	
	APLICOR 3D Printer Cleaning & Disinfection		
	a. Remove Dispenser 1 and 3D Regen PCL from APLICOR 3D Printer and dispose them in the clinical waste bin	1	
	b. Remove Bed Film from Matrix Plate and dispose it in the clinical waste bin	1	
11	c. Follow the user instructions of the disinfectant product to clean the inner chamber of APLICOR 3D Printer	1	
11	d. Turn on the UV lamp and disinfect for at least 30 minutes.	1	
	e. Turn off the UV lamp	1	
	f. Turn off APLICOR 3D Printer and unplug the power cable	1	
	g. Clean the used Matrix Plate and Sensor Tray	1	
	h. Sterilize Matrix Plate and Sensor Tray	1	